

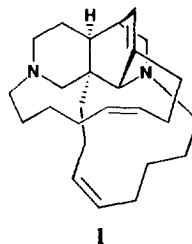
## Chiral Resolution of (±)-Keramaphidin B and Isolation of Manzamine L, a New β-Carboline Alkaloid from a Sponge *Amphimedon* sp.

Masashi Tsuda, Kenjiro Inaba, Naoko Kawasaki, Kaori Honma, and  
 Jun'ichi Kobayashi\*

Faculty of Pharmaceutical Sciences, Hokkaido University, Sapporo 060, Japan

**Abstract:** Both enantiomers of keramaphidin B (**1**) were separated by using chiral HPLC, of which one may be a plausible biogenetic precursor of ircinals A (**4**) and B (**5**) and manzamines A (**2**) and B (**3**), while the other may be associated with antipodes of manzamine alkaloids such as ircinols A (**6**) and B (**7**). Isolation and structure elucidation of a new manzamine congener, manzamine L (**8**), and absolute configurations at C-1 of **8** and manzamines D (**10**) and H (**9**) deduced from the Cotton effects are also described.

Keramaphidin B<sup>1</sup> (**1**) is a unique pentacyclic alkaloid with an unprecedented skeleton isolated from an Okinawan marine sponge *Amphimedon* sp., while several alkaloids with similar skeletons to that of **1** such as ingenamine,<sup>2</sup> ingenamines B ~ F,<sup>3</sup> ingamines A and B,<sup>4</sup> and xestocyclamines A<sup>5</sup> and B<sup>6</sup> from sponges of genus *Xestospongia* have been recently reported. These structures are very close to those of the corresponding biogenetic intermediate of manzamines A<sup>7</sup> (**2**) and B<sup>8</sup> (**3**) proposed by Baldwin and Whitehead,<sup>9</sup> in which a bis-3-alkyldihydropyridine macrocycle (**a**) may be converted through a Diels-Alder-type [4+2] intramolecular cycloaddition into a pentacyclic intermediate like **1**, which turns out manzamines A (**2**) and B (**3**) via a tetracyclic intermediate such as ircinals A (**4**) and B (**5**).<sup>10</sup> This might be followed by that manzamines A (**2**) and B (**3**) and keramaphidin B (**1**) possess the same absolute configurations, and indeed, ircinals A (**4**) and B (**5**) were found to have the same absolute configurations as those of manzamines A (**2**) and B (**3**). However, a small amount of crystals of keramaphidin B (**1**) employed for X-ray analysis was revealed to be racemic<sup>1</sup> though it possessed four asymmetric centers, and ircinols A (**6**) and B (**7**) were found to have opposite configurations concerning the four asymmetric centers of manzamines and ircinals.<sup>11</sup> On the other hand, ingenamine, ingamine A, and ingenamine E were reported to be antipodal to manzamines and ircinals.<sup>3</sup>



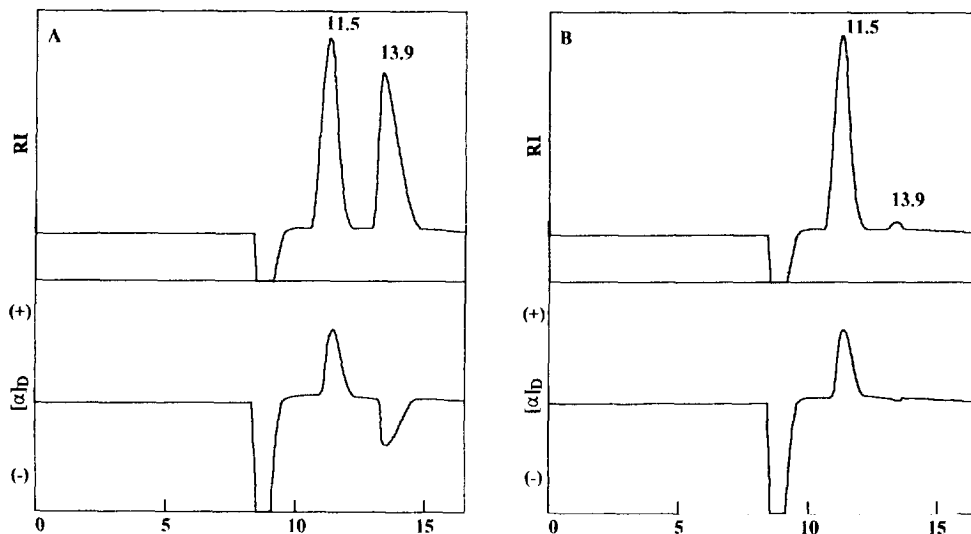
**1**

In this paper we describe the separation of (+)- and (-)-keramaphidin B (**1**) using chiral HPLC, the isolation and structure elucidation of a new manzamine congener, manzamine L (**8**), from a sponge *Amphimedon* sp., and a plausible biogenetic path of manzamines A (**1**) and B (**2**) on the basis of the structures and ratios of all the manzamine-related alkaloids isolated from this sponge.

EtOAc and *n*-BuOH soluble materials of MeOH extracts of the sponge *Amphimedon* sp. collected off Kerama Islands, Okinawa were purified by silica gel columns to give keramaphidins B (**1**, 0.0054 %, wet weight,  $[\alpha]_{\text{D}}^{20} +22.2^\circ$ ) and C<sup>12</sup> (0.0022 %), manzamines A (**2**, 0.081 %  $[\alpha]_{\text{D}}^{20} +46^\circ$ ), B (**3**, 0.0072 %,  $[\alpha]_{\text{D}}^{20} +93^\circ$ ), C<sup>8</sup> (0.002%), and D<sup>10,13</sup> (**10**, 0.012 %,  $[\alpha]_{\text{D}}^{24} +44^\circ$ ), ircinals A (**4**, 0.0009 %,  $[\alpha]_{\text{D}}^{15} +42^\circ$ ) and B (**5**, 0.0006 %,  $[\alpha]_{\text{D}}^{15} +15^\circ$ ), keramamine C<sup>12</sup> (0.0026 %), ircinols A (**6**, 0.006 %,  $[\alpha]_{\text{D}}^{18} -19^\circ$ ) and B (**7**, 0.0003 %,  $[\alpha]_{\text{D}}^{18} -2.8^\circ$ ), 6-hydroxymanzamine A<sup>14</sup> (= manzamine Y<sup>15</sup>, 0.005 %,  $[\alpha]_{\text{D}}^{20} +139^\circ$ ), 3,4-dihydrumanzamine A<sup>14</sup> (0.002 %,  $[\alpha]_{\text{D}}^{20} +86^\circ$ ), and 8-hydroxymanzamine A<sup>16</sup> (= manzamine G<sup>17</sup>, 0.002 %,  $[\alpha]_{\text{D}}^{20} +118^\circ$ ). Further separation of *n*-BuOH soluble materials with silica gel columns (CHCl<sub>3</sub>/MeOH and cyclohexane/acetone/Et<sub>2</sub>NH) and then silica gel HPLC (hexane/*i*-PrOH/Et<sub>3</sub>N) afforded manzamine L (**8**, 0.0056 %,  $[\alpha]_{\text{D}}^{24} -15^\circ$ ) together with manzamine H<sup>10</sup> (**9**, 0.0028 %,  $[\alpha]_{\text{D}}^{24} +21^\circ$ ).

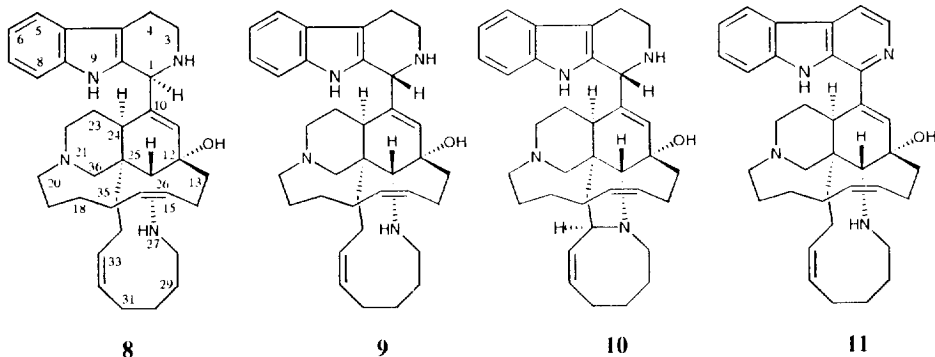
Keramaphidin B (**1**) was optically active as mentioned above, while a small amount of crystals of **1** obtained from CH<sub>3</sub>CN was racemic.<sup>1</sup> Chiral resolution of this racemate was performed by HPLC using a column [Chiralpak OP(+)] packed with (+)-poly(diphenyl-2-pyridylmethyl methacrylate) eluted with MeOH/H<sub>2</sub>O (80:20) (flow rate, 0.3 mL/min; RI and  $[\alpha]_{\text{D}}$  detection) and resulted in separation of (+)-keramaphidin B ((+)-**1**,  $t_{\text{R}}$  11.5 min) and (-)-keramaphidin B ((-)-**1**,  $t_{\text{R}}$  13.9 min) shown in Chart 1. Chiral HPLC analyses of the crystals and the mother liquid of keramaphidin B (**1**) revealed that the ratio of (+)- and (-)-forms of keramaphidin B (**1**) was ca. 1:1 and 20:1, respectively.

Manzamine L (**8**) showed the molecular ion peak at  $m/z$  554 ( $M^+$ ) in the EIMS spectrum, and



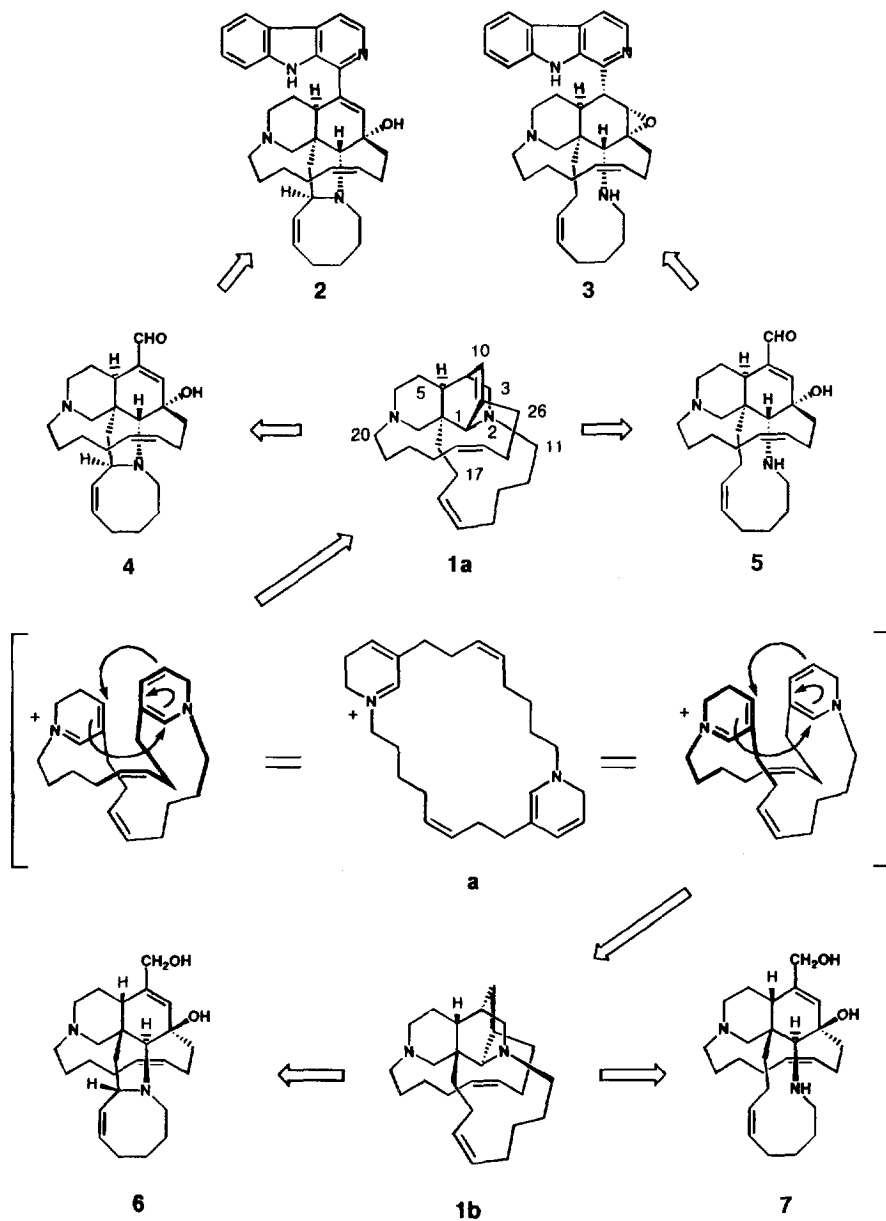
**Chart 1. Chiral HPLC of Keramaphidin B (**1**) in Resolution of Crystals (A) and Its Mother Liquid (B).**

Peaks:  $t_{\text{R}}$  11.5 min, (+)-Keramaphidin B {(+)-**1**};  $t_{\text{R}}$  13.9 min, (-)-Keramaphidin B {(-)-**1**}; HPLC Condition: eluent, MeOH/H<sub>2</sub>O (80:20); column, Chiralpak OP (+) (4.6 i.d. x 250mm); flow rate; 0.3 mL/min; RI and  $[\alpha]_{\text{D}}$  detection; sample injection, 50 $\mu$ g/100 $\mu$ L in MeOH.

Table 1.  $^{13}\text{C}$  NMR Data of Manzamines L (**8**) and H (**9**) in  $\text{CDCl}_3$ .

position	<b>8</b>		<b>9</b>		position	<b>8</b>		<b>9</b>	
1	56.1	d	59.9	d	17	28.1	t	29.1	t
3	41.3	t	43.2	t	18	27.5	t	28.6	t
4	21.7	t	22.4	t	19	28.2	t	29.2	t
4a	109.7	s	109.0	s	20	52.8	t	53.4	t
4b	126.6	s	127.8	s	22	48.6	t	49.6	t
5	117.1	d	118.0	d	23	31.7	t	32.3	t
6	118.6	d	119.3	d	24	46.2	d	44.6	d
7	120.6	d	121.4	d	25	42.9	s	43.5	s
8	110.0	d	111.0	d	26	58.1	d	59.2	d
8a	134.8	s	135.5	s	28	58.1	t	59.2	t
9a	132.6	s	134.2	s	29	28.2	t	29.2	t
10	143.3	s	143.9	s	30	28.2	t	29.2	t
11	128.6	d	130.1	d	31	24.1	t	25.0	t
12	68.9	s	70.1	s	32	130.6	d	131.6	d
13	39.6	t	40.6	t	33	130.0	d	131.1	d
14	21.0	t	21.9	t	34	25.1	t	26.2	t
15	128.4	d	129.4	d	35	37.0	t	37.3	t
16	127.92	d	129.1	d	36	64.9	t	65.7	t

HREIMS data ( $m/z$  554.3975,  $\text{M}^+$ ,  $\Delta$  -0.9 mmu) provided the molecular formula,  $\text{C}_{36}\text{H}_{50}\text{N}_4\text{O}$ , which was the same as that of manzamine H (**9**). The  $^1\text{H}$  NMR spectrum of **8** was similar to that of **9**, except for the signals due to H-1 ( $\delta$  4.57 for **8**;  $\delta$  4.65 for **9**) and H-11 ( $\delta$  5.36 for **8**;  $\delta$  5.64 for **9**). Comparison of the  $^{13}\text{C}$  NMR data (Table 1) of **8** with those of **9** indicated that manzamine L (**8**) was stereoisomer at C-1 of manzamine H (**9**); the carbon resonance at C-1 ( $\delta$  56.1) on the tetrahydro- $\beta$ -carboline ring in **8** slightly differed from that of **9** ( $\delta$  59.9). 1S-Configuration for manzamine L (**8**) was deduced from a negative Cotton effect ( $\Delta\epsilon$  -10.8) at 221 nm,<sup>18</sup> while manzamines H (**9**) and D<sup>19</sup> (**10**) isolated from this sponge showed the opposite sign ( $\Delta\epsilon$  +15.0 and +26.3, respectively) at 224 nm, implying 1R-configuration for **9** and **10**. The stereochemistry of manzamine L (**8**) was established by chemical correlation with manzamine J<sup>10</sup> (**11**) as follows. Treatment of **8** with 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) yielded the corresponding  $\beta$ -carboline (**11**), which was identical with manzamine J (**11**) in comparison with all spectral data of **11**, indicating that the isoquinoline moiety of manzamine L (**8**) possessed the same absolute stereochemistry as those of manzamines A (**2**), B (**3**), and J (**11**).



**Scheme 1. Plausible Biogenetic Path of Manzamines A (2) and B (3).**

One enantiomer of keramaphidin B (**1**) is considered to possess the same configuration as most of manzamine-related alkaloids with dextrorotation represented by manzamines A (**2**) and B (**3**), while the other enantiomer may be correlated to those with raevorotation like ircinols A (**6**) and B (**7**). The ratio of the total amount of manzamine-related alkaloids with dextrorotation vs. those with raevorotation in this sponge was found to be about 15:1, which approximately corresponded to the ratio of (+)- and (-)-keramaphidin B (**1**).

A plausible biogenetic path of manzamines A (**2**) and B (**3**) is shown in Scheme 1, which was elucidated on the basis of the structures and ratios of **1** ~ **10** isolated from this sponge. The Baldwin and Whitehead *bis*-3-alkyldihydropyridine (**a**) might be a biogenetic precursor of both enantiomers of keramaphidin B (**1**), and 2,3-iminium form of the major enantiomer (**1a**) may be hydrolyzed to generate (+)-ircinols A (**4**) or B (**5**), which are probably converted through Pictet-Spengler cyclization with tryptamine into manzamines D (**10**), H (**9**), and L (**8**), respectively, and then dehydrogenated into manzamines A (**2**) and B (**3**), respectively, while the minor enantiomer (**1b**) may be associated with some antipodes such as ircinols A (**6**) and B (**7**). The ratio (15:1) of amount of manzamine-related alkaloids with dextrorotation vs. those with raevorotation may indicate an incomplete stereoselectivity in *endo*-type Diels-Alder-type [4+2] intramolecular cycloaddition of the *bis*-3-alkyldihydropyridine (**a**) to afford a major and a minor enantiomers of keramaphidin B (**1a** and **1b**, respectively).

Recently keramaphidin B (**1**) was isolated from the sponge *Xestospongia ingens* together with ingenamine.<sup>3</sup> Since the optical rotation ( $[\alpha]_D^{+22.2^\circ}$ ) of **1** from *Amphimedon* sp. was close to that ( $[\alpha]_D^{+29.8^\circ}$ ) of **1** from *X. ingens*, both samples might possess the same absolute configurations. The absolute configuration of **1** from each sponge, however, remains to be determined.

Manzamine L (**8**) exhibited cytotoxicity against murine lymphoma L1210 cells and human epidermoid carcinoma KB cells (IC<sub>50</sub> 3.7 and 11.8 μg/mL, respectively) and antibacterial activity against bacteria, *Sarcina lutea*, *Staphylococcus aureus*, *Bacillus subtilis*, and *Mycobacterium* 607 (MIC 10, 10, 10, and 5 μg/mL, respectively).

## Experimental

**Isolation.** The sponge *Amphimedon* sp. (1.5 kg) collected off Kerama Islands, Okinawa, was extracted with MeOH (1 L x 2), and the extract was partitioned between EtOAc (300 mL x 3) and water (500 mL), and the aqueous layer was subsequently extracted with *n*-BuOH (300 mL x 3). A part (960 mg) of the *n*-BuOH soluble material (5.60 g) was chromatographed on silica gel columns (CHCl<sub>3</sub>/MeOH, 85:15 and then cyclohexane/acetone/Et<sub>2</sub>NH, 95:5:0.5) to give a residue (37 mg), a part of which (6.5 mg) was purified by silica gel column chromatography (*n*-hexane/isopropanol/Et<sub>3</sub>N, 95:5:0.5) to yield manzamine L (**8**, 4.2 mg, 0.0056 %, *t*<sub>R</sub> 12.8 min) together with manzamine H (**9**, 2.0 mg, 0.0028 % *t*<sub>R</sub> 14.9 min). Keramaphidins B (**1**, 0.0054 %, wet weight) and C<sup>12</sup> (0.0022 %), manzamines A (**2**, 0.081 %), B (**3**, 0.0072 %), C (0.002 %), and D (**10**, 0.012 %), ircinols A (**4**, 0.0009 %) and B (**5**, 0.0006 %), keramamine C (0.0026 %), ircinols A (**6**, 0.006 %) and B (**7**, 0.0003 %), 6-hydroxymanzamine A (0.005 %), 3,4-dihydromanzamine A (0.002 %), and 8-hydroxymanzamine A (0.002%) were isolated from other EtOAc and *n*-BuOH soluble fractions.

**Determination of the Ratio of (+)- and (-)-Keramaphidin B ((+)- and (-)-**1**).** The HPLC system consisted of a Chiralpak OP (+) column (4.6 i.d. x 250 mm, Daicel Chemical Ind., Ltd.), Shodex RI SE-61 and OP-1 detectors (Showa Denko Ltd.). A racemic form ((±)-**1**) of keramaphidin B crystallized from CH<sub>3</sub>CN was subjected to chiral HPLC analysis (eluent MeOH/H<sub>2</sub>O, 80:20; flow rate, 0.3 mL/min; sample injection, 50 μg/100 μL in MeOH) and peaks due to (+)- and (-)-forms were detected at *t*<sub>R</sub> 11.5 and 13.9 min in the ratio of 1:1, respectively. The mother liquid of keramaphidin B (**1**,  $[\alpha]_D^{+22.2^\circ}$ )

were also analyzed under the same condition as described above and the ratio of (+)- and (-)-forms of keramaphidin B (**1**) was estimated to be ca. 20:1.

**Manzamine L (8).** A colorless amorphous solid; mp 143 °C;  $[\alpha]_D^{24}$  -15° (*c* 0.42, CHCl<sub>3</sub>); IR (NaCl)  $\nu_{\max}$  3400 and 2990 cm<sup>-1</sup>; UV (MeOH)  $\lambda_{\max}$  223 ( $\epsilon$  34000) and 283 (6400) nm; CD (MeOH)  $\lambda_{\text{ext}}$  202 ( $\Delta\epsilon$  +13.6), 222 (-10.8), and 270 (+4.2) nm; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.82 (1H, brs), 7.48 (1H, d, *J* = 7.7 Hz), 7.31 (1H, d, *J* = 8.1 Hz), 7.14 (1H, t, *J* = 8.0 Hz), 7.09 (1H, t, *J* = 8.0 Hz), 5.64 (1H, s), 5.58 (1H, m), 5.43 (1H, m), 5.32 (1H, m), 5.19 (1H, m), 4.65 (1H, m), 3.74 (1H, brs), and 3.6 ~ 1.0 (36H, m); <sup>13</sup>C NMR (see Table 1); EIMS *m/z* 554 (M<sup>+</sup>); HREIMS *m/z* 554.3975 (M<sup>+</sup>, calcd for C<sub>36</sub>H<sub>50</sub>N<sub>4</sub>O, 554.3984).

**DDQ Oxidation of Manzamine L (8).** To a stirred solution of manzamine L (**8**, 5.0 mg) in EtOH (0.25 mL) and CHCl<sub>3</sub> (0.75 mL) at room temperature was added DDQ (3.3 mg). The mixture was stirred for 60 min. After evaporation of the solvent, the residue was purified by preparative silica gel TLC with CHCl<sub>3</sub>/MeOH (97:3) to afford Manzamine J (**11**, 2.0 mg): colorless amorphous solid; mp 138 ~ 139 °C;  $[\alpha]_D^{24}$  +45° (*c* 0.2, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  9.03 (1H, brs), 8.44 (1H, d, *J* = 5.4 Hz), 8.11 (1H, d, *J* = 7.8 Hz), 7.83 (1H, d, *J* = 5.4 Hz), 7.55 (2H, m), 7.26 (1H, m), 6.24 (1H, s), 5.64 (1H, m), 5.45 (1H, m), 5.35 (1H, m), 5.25 (1H, m), 3.90 (1H, s), and 3.32 (1H, m); EIMS *m/z* 550 (M<sup>+</sup>); HREIMS *m/z* 550.3696 (M<sup>+</sup>, calcd for C<sub>36</sub>H<sub>46</sub>N<sub>4</sub>O, 550.3672).

### Acknowledgement

We thank Prof. T. Sasaki, Kanazawa University, for the cytotoxicity test and Dr. Y. Mikami, Chiba University, for the antibacterial assay. This work was partly supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Science, Sport, and Culture of Japan.

### References and Notes

1. Kobayashi, J.; Tsuda, M.; Kawasaki, N.; Matsumoto, K.; Adachi, T. *Tetrahedron Lett.*, **1994**, *35*, 4383-4386.
2. Kong, F.; Andersen, R. J.; Allen, T. M. *Tetrahedron Lett.*, **1994**, *35*, 1643-1646.
3. Kong, F.; Andersen, R. J. *Tetrahedron*, **1995**, *51*, 2895-2906.
4. Kong, F.; Andersen, R. J.; Allen, T. M. *Tetrahedron*, **1994**, *50*, 6137-6144.
5. Roriguez, J.; Peters, B. M.; Kurz, L.; Schartzman, R. C.; McCarley, D.; Lou, L.; Crews, P. *J. Am. Chem. Soc.*, **1993**, *115*, 10436-10437.
6. Roriguez, J.; Crews, P. *Tetrahedron Lett.*, **1994**, *35*, 4719-4722.
7. (a) Sakai, R.; Higa, T.; Jefford, C. W.; Bernardinelli, G. *J. Am. Chem. Soc.*, **1986**, *108*, 6404-6405. (b) Nakamura, H.; Deng, S.; Kobayashi, J.; Ohizumi, Y.; Tomotake, Y.; Matsuzaki, T.; Hirata, Y. *Tetrahedron Lett.*, **1987**, *28*, 621-624.
8. Sakai, R.; Kohmoto, T.; Higa, T.; Jefford, C. W.; Bernardinelli, G. *Tetrahedron Lett.*, **1987**, *28*, 5493-5496.
9. Baldwin, J. E.; Whitehead, R. C. *Tetrahedron Lett.*, **1992**, *33*, 2059-2062.
10. Kondo, K.; Shigemori, H.; Kikuchi, Y.; Ishibashi, M.; Sasaki, T.; Kobayashi, J. *J. Org. Chem.*, **1992**, *57*, 2480-2483.
11. Tsuda, M.; Kawasaki, N.; Kobayashi, J. *Tetrahedron*, **1994**, *50*, 7957-7960.
12. Tsuda, M.; Kawasaki, N.; Kobayashi, J. *Tetrahedron Lett.*, **1994**, *35*, 4387-4388.
13. Higa, T. *Studies in Natural Product Chemistry B II*; Atta-ur-Rahman, Ed.; Elsevier: Amsterdam, 1989; pp346-353.
14. Kobayashi, J.; Tsuda, M.; Kawasaki, N.; Sasaki, T.; Mikami, Y. *J. Nat. Prod.*, **1994**, *57*, 1737-1740.
15. Kobayashi, M.; Chen, Y.-J.; Aoki, S.; In, Y.; Ishida, T.; Kitagawa, I. *Tetrahedron Lett.*, **1995**, *51*, 3727-3736.
16. Ichiba, T.; Corgiat, J. M.; Scheuer, P. J.; Kelly-Borges, M. *J. Nat. Prod.*, **1994**, *57*, 168-170.
17. Higa, T. *Proceedings of the First Princess Chulabhorn Science Congress 1987*; 1987; pp 450-459.
18. Stöckigt, J.; Zenk, M. H. *J. Chem. Soc., Chem. Commun.*, **1977**, 646-648.
19. 1*S*-Isomer of manzamine D has not been isolated from this sponge and the sponge of the genus *Ircinia*, from which ircinals A and B and manzamines H and J were isolated.